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Hyoliths are Palaeozoic lophophorates

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Hyoliths – orthothecids and hyolithids – are abundant and globally distributed ‘shelly’ fossils that appear early in the Cambrian and occur throughout the 280 million year span of Palaeozoic strata^{1,2}. The ecological and evolutionary significance of this group has remained unresolved, largely because of their idiosyncratic scleritome (operculum, conical shell, and paired ‘helens’ in hyolithids) and poorly constrained soft anatomy³⁻⁵. Since their first description over 175 years ago, hyoliths have most often been regarded as *incertae sedis*^{4,6}, allied with molluscs^{7,8} or assigned their own phylum^{1,2}. Here, we examine over 1500 specimens of the mid-Cambrian hyolithid *Haplophrentis* from the Burgess Shale and Spence Shale *Lagerstätten*. We reconstruct *Haplophrentis* as a semi-sessile epibenthic suspension feeder that was capable of using its helens to elevate its tubular body above the sea floor^{3,9-12}. Exceptionally preserved soft tissues include an extendable, gullwing-shaped, tentacle-bearing organ surrounding a central mouth, which we interpret as a lophophore, and a U-shaped digestive tract ending in a dorsolateral anus. Together with opposing bilateral sclerites and a deep ventral visceral cavity, these features indicate an affinity with the lophophorates (brachiopods, phoronids and tommotiids), substantially increasing the early disparity of this prominent group. This study reiterates the importance of soft-tissue preservation from Burgess Shale-type deposits in elucidating the evolutionary history of long-problematic taxa.

The radiation of biomineralized skeletons ranks among the most important and conspicuous records of the Cambrian Explosion. These ‘shelly’ Cambrian fossils track the evolution of animal body plans – but many have proved difficult to interpret from a biological perspective⁶. One enigmatic group, particularly widespread and diverse in Cambrian sediments, are the hyoliths. Orthothecid hyoliths possess a conical shell (previously referred to as “conch”) and cap-like operculum, each of which were putatively aragonitic and grew by basal-marginal accretion¹. Hyolithid hyoliths are additionally characterized by a pair of logarithmically curving lateral spines (‘helens’) and a non-planar operculum with a more extensive array of internally directed processes and muscle scars^{13,14}. Hyoliths have generally been considered as close allies of molluscs due to the presence of a bulb-shaped larval ‘protoconch’¹⁵ and purported mineralogical and microstructural similarities between their shells⁷ – though recent studies have questioned both the validity and the significance of these observations^{1,16}. Beyond this, the peculiar hyolith scleritome affords few opportunities for comparison with extant organisms. The only abundant evidence of non-mineralized anatomy comes from putative muscle scars, but these do little to constrain the affinities of hyoliths^{11,14}. Rarer traces of U-shaped guts³⁻⁵ are also phylogenetically uninformative due to their widespread occurrence among lophotrochozoans¹⁹.

Here, we revisit the systematic affinity of the hyolithid *Haplophrentis* based on over 1500 specimens, 254 of which preserve soft tissues (SI Discussion and Tables 1-2). The specimens include *Haplophrentis carinatus* from the Burgess Shale (principally from the Stanley Glacier¹⁷ and Marble Canyon¹⁸ localities, B.C., Canada) and *H. reesei*⁵ from the Spence Shale (Utah, U.S.A.), and are housed at the Royal Ontario Museum (ROM) and the Kansas University Museum of Invertebrate Paleontology (KUMIP), respectively.

Description. A gullwing-shaped band below the operculum emits as many as 12 (*H. carinatus*) to 16 (*H. reesei*) elongate elements that exhibit variable orientation and curvature (‘tentacles’ herein; Figs 1b, 2; Extended Data Figs 1-4). The lateral regions of the band may curve slightly posteriad or anteriad. The flared basal portion of each tentacle is followed by a longer, gently tapering section, up to half the length

of the operculum and about 7% as wide as long. Two shorter medial tentacles attach proximally around a centrally located mouth. We interpret this tentaculate band as a lophophore.

The lophophore connects to a wide medially situated structure that splays anteriorly from a narrow central tube with the mouth at the distal end. We interpret this tube as a pharynx with muscular walls. Variations in the length and width of this pharyngeal organ indicate that it, along with the attached lophophore, was protrusible. In some specimens, the pharyngeal organ is less than half the length of the operculum, and the distally attached tentacles are almost entirely concealed beneath the operculum (Fig. 2a-b; Extended Data Fig. 1). In others, the pharynx extends from the posterior of the operculum almost to its anterior margin, resulting in the tentacles projecting through the commissure (Figs 1b, 2c-d; Extended Data Figs 2-4). The aboral end of the pharyngeal organ was evidently attached to the operculum, as the pharynx and tentacles remain medially located when the operculum is displaced from the conical shell aperture (Fig. 1b; Extended Data Figs 1-4).

The pharynx narrows as it passes under the posterior margin of the operculum, joining with the gut (Fig. 1b; Extended Data Fig. 5). The undifferentiated, U-shaped gut is contained within the conical shell. It extends to at most 75% of the depth of the conical shell before looping back along the functional dorsum (Fig. 1b-c; Extended Data Fig. 5). The anus opens near the commissure, slightly left of the midline³ and outside the crown of tentacles (Fig. 1c; Extended Data Fig. 5a-b).

Paired, kidney-shaped traces in the carbon film, often surrounded by dark, carbonaceous rims, occur below the operculum, dorsal to the pharynx (Fig. 2a, c; Extended Data Figs 1a, b, 3, 6f). A dark, radial structure of equivalent constitution occurs below the umbo of the operculum, (Fig. 2a, c; Extended Data Figs 1a-b, 3, 6d-f). Muscle scars occupy a similar position in other hyolithids¹⁴, suggesting that the surrounding carbon is preserved connective tissue.

A symmetrical pair of large grey structures flanks the gut, along the functional dorsum of the conical shell, terminating near the posterior margin of the operculum (Fig. 1; Extended Data Figs 2c, 3b, 5a, 6a-c,

7b). At this point, each structure is nearly half the width of the operculum. The structures taper apically and – decay notwithstanding – extend as far as the loop of the gut. The biological identity of these visceral organs is unclear.

A thin and deformable layer, possibly the body wall, circumscribes the inside of the conical shell. It stops short of the apex of the conical shell, apparently leaving the apical region unoccupied (Extended Data Fig. 1c, 6c).

The helens of *Haplophrentis* emerge at a slight downward angle from a lateral notch at the commissure of the conical shell and operculum, growing as open logarithmic spirals that twist helically along their length and pointing either anteriad or posteriad^{10,11,14,20} (Extended Data Fig. 7). Brachiopods are attached to the helens of four *Haplophrentis* specimens that have tightly articulated skeletons and preserved soft tissues: strong evidence that these associations occurred when both animals were alive (Extended Data Fig. 8). As such, the helens could not have been submerged below the sediment-water interface, and “retracted” helens observed within conical shells⁵ are likely a taphonomic artefact.

Discussion. The lophophore of *Haplophrentis* resembles the characteristic tentacular feeding organs of several suspension feeding taxa, particularly Entoprocta and Lophophorata (i.e. Brachiopoda and Phoronida, possibly including Ectoprocta)²¹. This indicates that *Haplophrentis* was a benthic suspension feeder^{3,9,12} rather than a deposit feeder^{5,22}. The downward-directed helens^{10,14} may have been rotated to elevate the commissure from the sediment-water interface (Fig. 3, Extended Data Fig. 7, SI Discussion), consistent with observations of encrusting organisms on both the dorsal and ventral surfaces of hyolithid conical shells⁹.

The disposition of the mouth and anus at the anterior and posterior margins of the hyolithid commissure indicates that the operculum and conical shell were dorsoventrally opposed – contrasting with molluscs, which typically secrete only dorsal skeletal elements (shells, opercula)¹⁰. As in brachiopods, the lophophore is contained in the mantle cavity formed by dorsal and ventral bilateral sclerites²³. The

combination of characters present in *Haplophrentis* and the lack of molluscan apomorphies³ decisively supports an affinity with the lophophorates, particularly the brachiopods.

The simple arrangement of tentacles in the *Haplophrentis* lophophore is strikingly similar to that in brachiopod larvae (Extended Data Figs 1d, 2e), but distinct from the complex lophophore arrangements that characterise most adult brachiopods²³. Neither does the *Haplophrentis* lophophore form the closed loop characteristic of crown-group brachiopods²³, instead diverging laterally in a manner that recalls the primitive phosphatic-shelled brachiopod *Heliomedusa orientalis*²⁴.

The partial attachment of the lophophore to the hyolithid operculum supports a homology with the dorsal (brachial) valve of brachiopods²³, identifying the opposing conical shell as ventral and potentially homologous with the brachiopod pedicle valve (Fig. 4). Although opposing valves are common to both hyolithids and brachiopods, the deep extension of the hyolithid visceral area into the conical shell differs markedly from the condition in most crown-group brachiopods, where the viscera are reduced to the space between the valves. This arrangement is found in some fossil taxa, notably lingulellotretids, which are close to the linguliform members of the brachiopod crown group²⁵; *Yuganotheca*, which is thought to represent an intermediate between phoronid and the brachiopod body plans²⁶; and *Lingulosacculus*, whose phylogenetic position is ambiguous²⁷. Hyolith shell microstructure lacks an obvious equivalent amongst brachiopods, though shell penetrating canals of similar size and preservation in both hyoliths¹ and obolellids (putatively basal calcareous brachiopods)²⁸ potentially support comparable skeletal secretion in these groups.

While a position within total group Lophophorata is well supported, the ultimate placement of hyolithids will depend on the order of character acquisition in the brachiopod body plan. The dominant viewpoint suggests that brachiopods diverged from among the problematic tommotiids^{29,30}, and as such their ancestral ground-plan is inferred to have included a multi-element phosphatic scleritome. Working under this hypothesis, the conical shell, operculum and helens of hyolithids might be homologised with the sclerites of a *Micrina*-like tommotiid, resolving hyolithids (potentially including *Lingulosacculus*) in the

brachiopod stem lineage (Fig. 4). *Yuganotheca* – in which the lophophore is enclosed by unmineralized mantle lobes that protrude above a single ventral sclerite²⁶ – could then be interpreted as having undergone a secondary loss of mineralization. The ventrally extended viscera of these taxa conceivably characterise the ancestral form of crown group Brachiopoda, providing a link with the phoronid body plan²⁶. Under this interpretation, the elongated visceral cavity of lingulellotretids²⁵ may have been retained from the ancestral brachiopod state, with the more restricted visceral area of modern brachiopods arising once in linguliforms and again in rhynchonelliforms.

This hypothesis must remain somewhat speculative and incomplete, not least because a detailed comparison of hyolithids and tommotiids is made difficult by differences in sclerite mineralogy and microstructure^{1,30} (Extended Data 9-10, SI Discussion). Similar issues have frustrated attempts to interpret the evolution of morphologically distinct phosphatic- and calcitic-shelled brachiopod lineages²⁸. The addition of purportedly aragonitic hyoliths to the lophophorate tree further complicates the evolution of biomineralization in this group, emphasising the many details of deep lophotrochozoan phylogeny that remain ambiguous. Even so, the recognition of hyoliths as members of this clade settles a longstanding palaeontological debate, and emphasizes the high level of disparity and ecological dominance achieved by lophophorates in marine communities throughout the Palaeozoic Era.

Methods

The fossil material studied herein is deposited at the Royal Ontario Museum, Toronto (ROM) and the Kansas Museum of Invertebrate Paleontology, Kansas (KUMIP). Some ROM specimens were mechanically prepared using a tungsten-tipped micro-engraving tool. Specimens were photographed under various lighting conditions and with ammonium chloride coating to enhance contrast, and imaged using backscatter and secondary electron microscopy. Measurements were taken using ImageJ.

Data availability statement. Data generated or analysed during this study are included in this published article (and its supplementary information files).

149 **References**

- 150 1. Kouchinsky, A. V. Skeletal microstructures of hyoliths from the early Cambrian of Siberia.
 151 *Alcheringa*. **24**, 65-81 (2000).
- 152 2. Runnegar, B. et al. Biology of the Hyolitha. *Lethaia*. **8**, 181-191 (1975).
- 153 3. Martí Mus, M. A hyolithid with preserved soft parts from the Ordovician Fezouata *Konservat-*
 154 *Lagerstätte* of Morocco. *Palaeogeography, Palaeoclimatology, Palaeoecology*. **460**, 122-129 (2016).
- 155 4. Devaere, L., Clausen, S., Álvaro, J. J., Peel, J. S., Vachard, D. Terrenewian orthothecid (Hyolitha)
 156 digestive tracts from northern Montagne Noire, France; taphonomic, ontogenetic and phylogenetic
 157 implications. *Plos One*. **9**, e88583 (2014).
- 158 5. Babcock, L. E. & Robison, R. A. Taxonomy and paleobiology of some middle Cambrian *Scenella*
 159 (Cnidaria) and hyolithids (Mollusca) from western North America. *The University of Kansas*
 160 *Paleontological Contributions*. **Paper 121**, 1-22 (1988).
- 161 6. Bengtson, S., Conway Morris, S. Cooper, B., Jell, P. & Runnegar, B. Early Cambrian fossils from
 162 South Australia. *Association of Australasian Palaeontologists*, **Memoir 9**, 1-364 (1990).
- 163 7. Malinky, J. M. & Yochelson, E. L. On the systematic position of the Hyolitha (Kingdom Animalia).
 164 *Memoirs of the Association of Australasian Palaeontologists*. **34**, 521-536 (2007).
- 165 8. Marek, L. & Yochelson, E. L. Aspects of the biology of Hyolitha (Mollusca). *Lethaia*. **9**, 65-84
 166 (1976).
- 167 9. Galle, A. & Parsley, R. L. Epibiont relationships on hyolithids demonstrated by Ordovician
 168 trepostomes (Bryozoa) and Devonian tabulates (Anthozoa). *Bulletin of Geosciences*. **80**, 125-138
 169 (2005).
- 170 10. Runnegar, B. Hyolitha: status of the phylum. *Lethaia*. **13**, 21-25 (1980).

- 171 11. Martí Mus, M., Jeppsson, L. & Malinky, J. M. A complete reconstruction of the hyolithid skeleton.
172 *Journal of Paleontology*. **88**, 160-170 (2014).
- 173 12. Marek, L., Parsley, R. L. & Galle, A. Functional morphology of hyolithids based on flume studies.
174 *Věstník Českého geologického ústavu*. **72**, 351-358 (1997).
- 175 13. Martí Mus, M. & Bergström, J. Skeletal microstructure of helens, lateral spines of hyolithids.
176 *Palaeontology*. **50**, 1231-1243 (2007).
- 177 14. Martí Mus, M. & Bergström, J. The morphology of hyolithids and its functional implications.
178 *Palaeontology*. **48**, 1139-1167 (2005).
- 179 15. Dzik, J. Larval development of hyolithids. *Lethaia*. **11**, 293-299 (1978).
- 180 16. Skovsted, C. B. et al. The operculum and mode of life of the lower Cambrian hyolith *Cupithea* from
181 South Australia and north China. 2016, *Palaeogeography, Palaeoclimatology, Palaeoecology*. **443**,
182 123-130 (2016).
- 183 17. Caron, J.-B., Gaines, R., Mángano, G., Streng, M. & Daley, A. A new Burgess Shale-type
184 assemblage from the "thin" Stephen Formation of the Southern Canadian Rockies. *Geology* **38**, 811-
185 814 (2010).
- 186 18. Caron, J.-B., Gaines, R. R., Aria, C., Mángano, M. G. & Streng, M. A new phyllopod bed-like
187 assemblage from the Burgess Shale of the Canadian Rockies. *Nature Communications* **5**, 3210
188 doi:10.1038/ncomms4210 (2014).
- 189 19. Budd, G. E. & Jackson, I. S. C. Ecological Innovations in the Cambrian and the origin of crown
190 group phyla. *Philosophical Transactions of the Royal Society B*. **371**, 20150287 (2016).
- 191 20. Butterfield, N. J. and Nicholas, C. J. Burgess Shale-type preservation of both non-mineralizing and
192 'shelly' Cambrian organisms from the Mackenzie Mountains, northwestern Canada. *Journal of*
193 *Paleontology*. **70**, 893-899 (1996).
- 194 21. Kocot, K. M. On 20 years of Lophotrochozoa. *Organisms Diversity & Evolution*. **16**: 329-343 (2016).

22. Sun, H., Babcock L. E., Peng, J. & Zhao, Y. Three-dimensionally preserved digestive systems of two Cambrian hyolithides (Hyolitha). *Bulletin of Geosciences*. **91**, 51-56 (2016).
23. Kuzmina, T. V. & Malakhov, V. V. Structure of the brachiopod lophophore. *Paleontological Journal*. **41**, 520-536 (2007).
24. Zhang, Z. et al. Architecture and function of the lophophore in the problematic brachiopod *Heliomedusa orientalis* (early Cambrian, south China). 2009, *Geobios*. **42**, 649-661 (2009).
25. Zhang, Z. et al. Note on the gut preserved in the Lower Cambrian *Lingulellotreta* (Lingulata, Brachiopoda) from southern China. *Acta Zoologica*. **88**, 65-70 (2007).
26. Zhang, Z. et al. An early Cambrian agglutinated tubular lophophorate with brachiopod characters. *Scientific Reports*. **4**, 4682 (2014).
27. Balthasar, U. & Butterfield, N. J. Early Cambrian "soft-shelled" brachiopods as possible stem-group phoronids. *Acta Palaeontologica Polonica*. **54**, 307-314 (2009).
28. Balthasar, U. *Mummpikia* gen. nov. and the origin of calcitic-shelled brachiopods. *Palaeontology*. **51**, 263-279 (2008).
29. Skovsted, C. B., Brock, G. A., Paterson, J. R., Holmer, L. E. & Budd, G. E. The scleritome of *Eccentrotheca* from the Lower Cambrian of South Australia: Lophophorate affinities and implications for tommotiid phylogeny. *Geology*. **36**, 171-174 (2008).
30. Murdock, D. J. E., Bengtson, S., Marone, F., Greenwood, J. M. & Donoghue, P. C. J. Evaluating scenarios for the evolutionary assembly of the brachiopod body plan. *Evolution & Development*. **16**, 13-24 (2014).

Supplementary Information is available in the online version of the paper.

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Figures

Figure 1 | *Haplophrentis carinatus* from the Burgess Shale. Specimens oriented anterior to the top, fossil images (top row) and associated line drawings (bottom row). **a**, Royal Ontario Museum (ROM) 62928.5 from Marble Canyon, dorsal view with partially broken operculum, showing the gut and paired visceral organs within the conical shell. **b**, ROM63981.1 from Stanley Glacier, dorsal view (composite image of part and counterpart) showing the lophophore attached to the pharynx and most of the gut. **c**, ventral view of ROM63982.1 from Stanley Glacier, showing the U-shaped gut and anus. Scale bars = 2 mm. Abbreviations: a, anus; ag, anal branch of gut; c, conical shell; cl, clavicle; cp, cardinal process; lh, left helen; m, mouth; o, operculum; og, oral branch of gut; p, pharynx; rh, right helen; t, tentacle; vo, visceral organ.

Figure 2 | Soft tissues associated with the *Haplophrentis* operculum. Dorsal view of specimens, anterior to the top. **a**, ROM63983.1, *H. carinatus* from Stanley Glacier, lophophore in retracted position with at least 6 tentacles on the left side, showing muscle scars. **b**, Kansas Museum of Invertebrate

Paleontology (KUMIP) 366447, *H. reesei* from the Spence Shale, showing retracted lophophore with 16 tentacles. **c**, ROM59943.1, *H. carinatus* from Stanley Glacier, showing partially extended lophophore with tentacles beyond the operculum margin. **d**, KUMIP 204340, *H. reesei* from the Spence Shale, with a fully extended pharynx and lophophore. Scale bars = 2 mm. Abbreviations: ct, connective tissue; g, gut; ms, muscle scar; mt, medial tentacle; pl, pharynx lumen; other abbreviations as in Figure 1.

Figure 3 | Anatomical reconstruction of *Haplophrentis*. a-d, lophophore extended, helens in anterior position. **a**, dorsal profile; **b**, left lateral profile; **c**, ventral profile, ligula of conical shell cut away; **d**, frontal profile, operculum removed. **e-f**, lophophore retracted, helens in posteriad (resting) position. **e**, left lateral profile; **f**, ventral profile, ligula of conical shell cut away. **g**, life reconstruction on the Cambrian sea floor. Abbreviations: bw, body wall; other abbreviations as in Figures 1, 2.

Figure 4 | Possible position of hyolithids within total group Lophophorata. Dashed lines indicate hypothesized lophophorate relationships. The inclusion of Ectoprocta within Lophophorata remains ambiguous²¹, hence they are omitted from this figure. 1 = lophophore, U-shaped gut curving ventrally, multielement phosphatic scleritome; 2 = dorsal and ventral valves enclosing lophophore chamber; 3 = aragonitic(?) sclerite mineralogy; 4 = pedicle with coelomic cavity, lateral elongation of lophophore arms; 5 = calcitic shell mineralogy, extreme reduction of visceral area, loss of anus. Colour scheme of diagrams: pink, visceral area; green, lophophore; purple, gut; blue, ventral valve; yellow, dorsal valve. Extinct taxa marked by a cross.

Extended Data Figure 1 | Retracted lophophore of *Haplophrentis*. a, b, *H. carinatus*, dorsal view of ROM63983.1 from Stanley Glacier. **a**, entire specimen photographed dry with polarized light; **b** detail of tissue associated with the operculum. **c**, dorsal view of KUMIP366447 from the Spence Shale photographed wet with polarized light. **d**, Larva of the extant brachiopod *Glottidia* with retracted lophophore; image reprinted from fig.1b in Strathmann, R. Ciliary sieving and active ciliary response in capture of particles by suspension-feeding brachiopod larvae. Acta Zoologica. Wiley. © 2005 The Royal Swedish Academy of Sciences. Scale bars: **a-c**, 2 mm; **d**, 0.2 mm. Abbreviations: bw, body wall; c,

270 conical shell; cl, clavicle; cp, cardinal process; ct, connective tissue; es, embryonic shell; g, gut; lh, left
271 helen; ls, larval shell; ms, muscle scar; mt, medial tentacle; o, operculum; pd, pedicle; rh, right helen; t,
272 tentacle.

273 **Extended Data Figure 2 | *Haplophrentis carinatus* from Stanley Glacier (ROM63981.1).** **a**,
274 operculum, showing extended pharynx and lophophore photographed dry with polarized light. **b**,
275 interpretive drawing. **c**, part, photographed wet with polarized light; **d**, counterpart, photographed wet
276 with polarized light. **e**, larva of the extant brachiopod *Glottidia* with extended lophophore; image
277 reprinted from fig. 1a in Strathmann, R. Ciliary sieving and active ciliary response in capture of particles
278 by suspension-feeding brachiopod larvae. Acta Zoologica. Wiley. © 2005 The Royal Swedish Academy
279 of Sciences. Scale bars: **a-d**, 2 mm; **e**, 0.2 mm. Abbreviations: ag, anal branch of gut; bw, body wall; cl,
280 clavicle; cp, cardinal process; m, mouth; og, oral branch of gut; p, pharynx; pl, pharynx lumen; t, tentacle.

281 **Extended Data Figure 3 | *Haplophrentis carinatus* from Stanley Glacier (ROM 59943.1).** **a**, part
282 photographed dry with polarized light; **b**, counterpart, photographed wet with polarized light. **c**,
283 operculum (composite image of part and counterpart) showing extended tentacles, photographed dry with
284 polarized light. Scale bars = 2 mm. Abbreviations: cl, clavicle; cp, cardinal process; ct, connective tissue;
285 g, gut; ms, muscle scar; pl, pharynx lumen; t, tentacle; vo, visceral organ.

286 **Extended Data Figure 4 | *Haplophrentis reesei* from the Spence Shale (KUMIP204340).** **a-b**,
287 operculum, showing extended pharynx and lophophore. **a**, photographed dry with polarized light. **b**, wet,
288 with polarized light. **c-e**, whole specimen. **c**, dry, unpolarized light. **d**, dry with polarized light. **e**, wet with
289 polarized light. Scale bars: **a-b**, 2 mm; **c-e**, 5 mm. Abbreviations: c, conical shell; ct, connective tissue; g,
290 gut; lh, left helen; m, mouth; o, operculum; p, pharynx; pl, pharynx lumen; rh, right helen; t, tentacle.

291 **Extended Data Figure 5 | U-shaped digestive tract of *Haplophrentis carinatus*.** **a, b**, ROM63982.1
292 from Stanley Glacier, ventral view, photographed wet with polarized light; **b** corresponds to area boxed in
293 **a**. **c**, ROM63984.1, dorsal view, photographed dry with polarized light. Scale bars = 1 mm.

Abbreviations: a, anus; ag, anal branch of gut; og, oral branch of gut; p, pharynx; t, tentacle; vo, visceral organ.

Extended Data Figure 6 | Musculature and visceral area of *Haplophrentis carinatus*. **a**, ROM63985.1 from Marble Canyon, laterally oriented specimen showing the position of visceral organs and gut within the conical shell, photographed wet with polarized light. **b**, ROM62928.5 from Marble Canyon, dorsal view showing paired visceral organs flanking the gut, photographed wet with polarized light. **c**, ROM63986.1 from Marble Canyon, dorsal view with paired visceral organs adjacent to the gut, photographed dry with polarized light. **d, e**, ROM63987.1 from Mount Odaray, photographed wet with polarized light. **d**, ventral view of the operculum, showing connective tissue dorsal to the pharynx; **e**, detail of area boxed in **d**. **f**, ROM63988.1 from Stanley Glacier, dorsal view of operculum with preserved muscle scars and connective tissue, dorsal to the pharynx, photographed dry with polarized light. Scale bars = 1 mm. Abbreviations: bw, body wall; ct, connective tissue; g, gut; m, mouth; ms, muscle scar; og, oral branch of gut; p, pharynx; t, tentacle; vo, visceral organ.

Extended Data Figure 7 | *Haplophrentis scleritome*. **a**, ROM62968.4 from Marble Canyon, lateral view; note downward disposition of right helens, which emerges from the commissure just above the ligula of the conical shell, photographed dry with polarized light. **b**, ROM62968.2, obliquely preserved specimen with anteriorly directed helens, showing the shape of the aperture of the conical shell, photographed dry with polarized light. **c**, Backscatter Scanning Electron Microscope image showing the bulb-shaped larval shell at apex of conical shell in ROM63989.1 from Marble Canyon. **d**, ROM63991.1 from Marble Canyon, with a slightly displaced operculum and the helens directed anteriorly and curving below the body, photographed dry with polarized light. **e**, ROM63993.1 from Marble Canyon, operculum closing the conical shell aperture, both helens directed posteriorly with the left one preserved in the same plane as the body and the right one curving below, photographed dry with polarized light. **f**, two specimens (ventral views) from Marble Canyon showing variation in the curvature and twist of the helens (visible portion is in the same plane as the conical shell in both), photographed wet with polarized light.

319 **f₁**, ROM64005.1; **f₂**, ROM63989.1. **g**, dorsal view of the right helen of ROM63992.1 from the Raymond
320 Quarry, curving posteriad (inserting into the body on the upper right side) with the direction of twist
321 indicated by the arrow; photographed using unpolarized light. **h**, ROM63994.1 from the Walcott Quarry,
322 backscatter SEM image of a helen showing ornament of transverse ribs. **i**, **j**, ROM63995.4 from the
323 Walcott Quarry, photographed wet with polarized light. **i**, whole specimen; **j**, detail of area boxed in **i**,
324 ornament of transverse ribs on the conical shell. Scale bars: **a**, **b**, **d**, **e**, **g**, **i**, 2 mm; **c**, 0.5 mm; **f₁**, **f₂**, **h**, **j**, 1
325 mm. Abbreviations: c, conical shell; cp, cardinal process; g, gut; lh, left helen; o, operculum; p, pharynx;
326 rh, right helen; vo, visceral organ.

327 **Extended Data Figure 8 | Brachiopod epibionts on *Haplophrentis***. Arrows indicate brachiopods. **a**,
328 ROM63996.1, *H. carinatus* with *Nisusia? burgessensis*, photographed using ammonium chloride
329 sublimate. **b-c**, ROM63997.1, *H. carinatus* with an acrotretid brachiopod, note soft tissue preserved
330 below operculum, photographed dry with polarized (**b**) and unpolarized light (**c**). **d-f**, KUMIP314211, *H.*
331 *reesei* with *Micromitra* sp., photographed using unpolarized light. **g-h**, KUMIP304352, *H. reesei* with
332 *Nisusia* sp., photographed using unpolarized light. Scale bars: **a-b**, 2 mm; **c**, **e**, **f**, **h**, 1 mm; **d**, **g**, 5 mm.

333 **Extended Data Figure 9 | Elemental distribution in *Haplophrentis carinatus* from Marble Canyon**
334 **(ROM63998.1)**. Scale bars = 2 mm. Abbreviations: PL, polarized light (photographed wet); SE,
335 secondary electron micrograph; C, carbon; S, sulfur; Mg, magnesium; Fe, iron; K, potassium; P,
336 phosphorous; Ca, calcium; Al, aluminum; Na, sodium; O, oxygen; Si, silicon; Ti, titanium.

337 **Extended Data Figure 10 | Detail of elemental composition of *Haplophrentis carinatus* from Marble**
338 **Canyon. a-d**, ROM63998.1. **a**, **b**, carbon maps of part and counterpart; note the concentration of carbon
339 in the transverse shell ornament, clavicles and cardinal processes – evidence of an organic component of
340 the skeleton; **c**, sulfur map (composite image of part and counterpart), showing, soft tissues, including
341 tentacles, partially replaced by pyrite. **d**, phosphorous map (composite image of part and counterpart),
342 showing phosphatized gut. **e-f**, ROM63999.1, carbon maps of part and counterpart, note that carbon
343 surrounding the clavicles and cardinal processes may be related to the attachment of muscles and

344 connective tissue in these regions. Scale bars = 1 mm. Abbreviations: cl, clavicle; cp, cardinal process; g,
345 gut; lh, left helen; p, pharynx; t, tentacle; vo, visceral organ.

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